BIO14199 : CRISPR barcoding: method and kit for labeling and detecting a population of endonucleasetreated cells

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BIO14199

CRISPR barcoding: method and kit for labeling and detecting a population of endonuclease-treated cells

Product

Product:

CRISPR-barcoding, a fast and highly flexible strategy which enables detection of cells containing the mutation of interest within a mass population of unmodified cells using real-time quantitative PCR or deep sequencing.

Application:

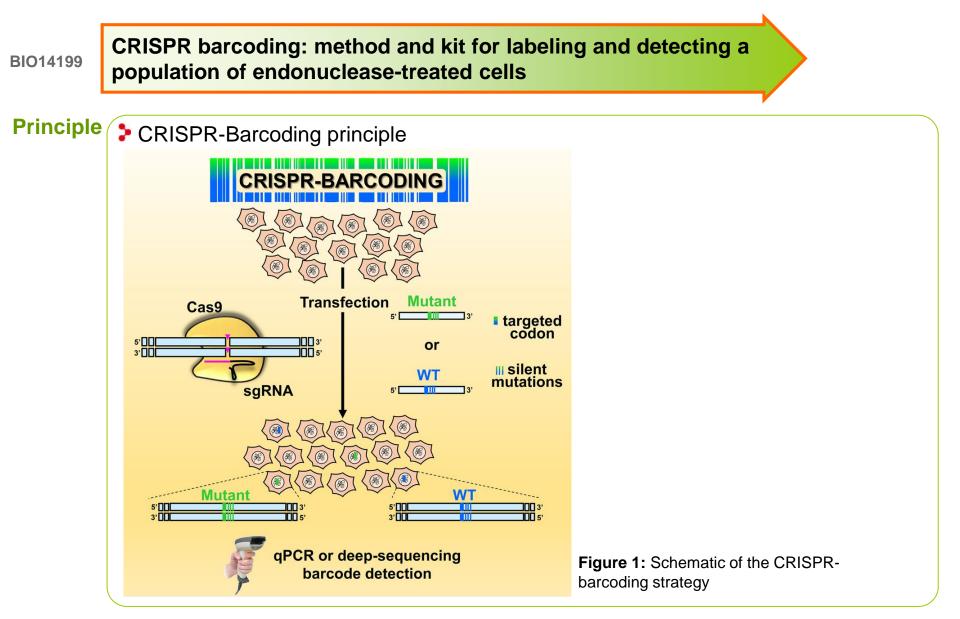
- alternative tool to the classical lentiviral DNA barcode libraries, ensuring the detection of thousands of distinct barcodes through qPCR or deep-sequencing.
- tracing of the mutated cells immediately after DNA editing without the need to derive clones, thus providing a unique means to investigate the effects of different kinds of genomic modifications, regardless of their potential impact on cell growth, in a broad range of functional assays.
- high-resolution tracking of single specific cancer cells allowing to identify even rare pre-existing resistant subclones potentially involved in mechanisms of acquired resistance to therapy.

Proof of concept:

The *in vitro* results were confirmed *in vivo*, using a CRISPR-barcoding xenograft model for NSCLC. The main proofs of concept reported are related to cancer models, nevertheless this technology can be implemented in different fields of biological research.

Patent and publication:

WO2017068120 (A1): Endonuclease-Barcoding



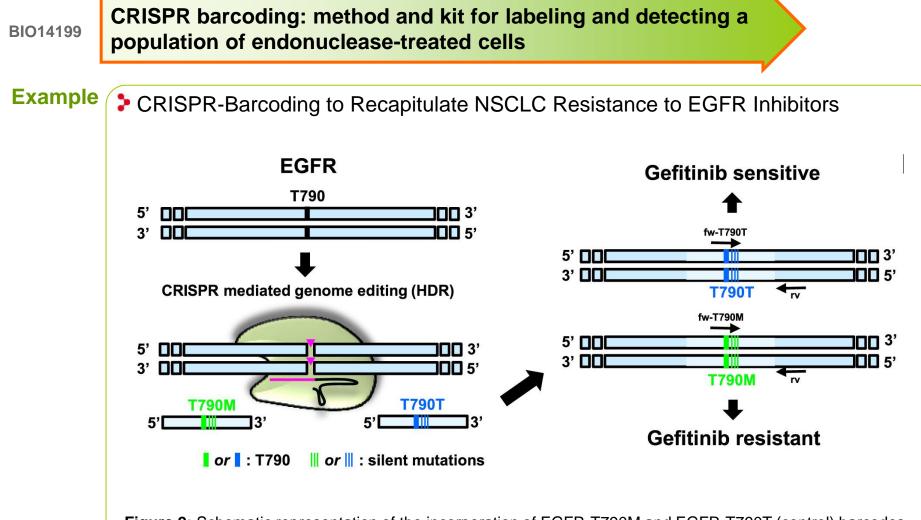


Figure 2: Schematic representation of the incorporation of EGFR-T790M and EGFR-T790T (control) barcodes in NSCLC cells.

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Example CRISPR-barcoding to functionally characterize oncogenic mutations in a context of intratumor heterogeneity.

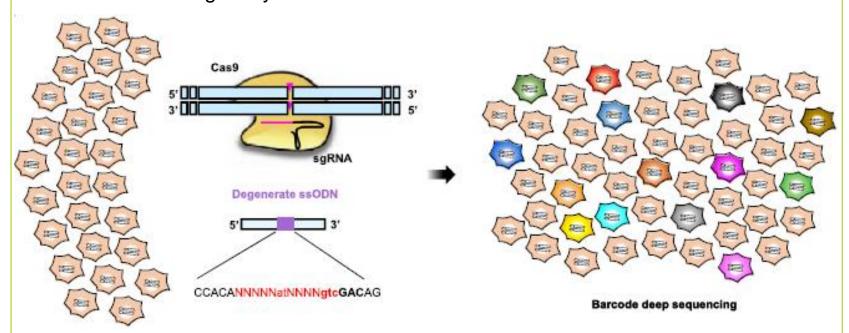


Figure 3: Through insertion of a highly complex series of degenerate sequences at a specific genomic location, CRISPR-barcoding can be used to trace several thousands of genetically labelled clones within a mass population of tumor cells.

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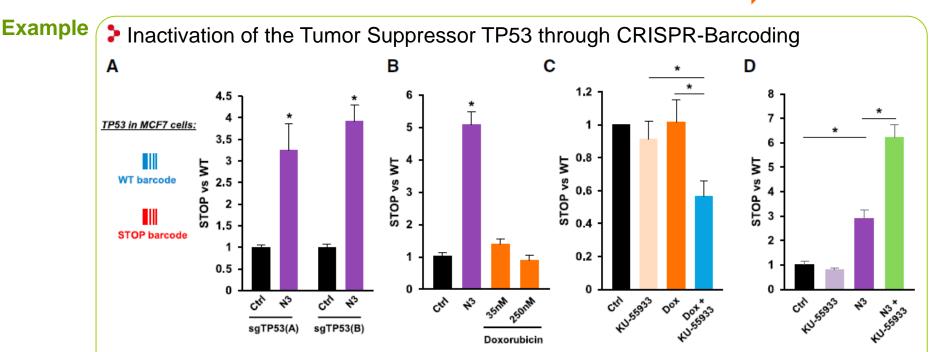


Figure 4:

(A) Effects of Nutlin-3 (N3; 10 mM, 7 days) on the TP53-STOP to TP53-WT ratio in MCF7 cells using two distinct sgRNAs (A or B). Mean \pm SEM; n = 4 of one representative of three independent experiments. *p < 0.05 (Mann-Whitney test).

(B) Effects of N3 (10 mM; 7 days) or doxorubicin (Dox) on the TP53-STOP to TP53-WT ratio in HCT-116 cells. Mean \pm SEM; n = 4 of one representative of three independent experiments.

(C) The cells in (B) were treated for 7 days with KU-55933 (10 mM) and/or Dox (50 nM), and the TP53-STOP to TP53-WT ratio was assessed by qPCR. Mean ± SEM of seven independent experiments.

(D) Effects of KU-55933 (10 mM) and/or N3 (10 mM, 7 days) on the TP53-STOP to TP53-WT ratio in HCT-116 cells. Mean ± SEM; n = 4 of one representative of three independent experiments.

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